# Package 'garfield'

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Type Package

**Title** GWAS Analysis of Regulatory or Functional Information Enrichment with LD correction

**Version** 1.32.0 **Date** 2015-12-14

Author Sandro Morganella <sm22@sanger.ac.uk>
Maintainer Valentina Iotchkova <vi1@sanger.ac.uk>

**Description** GARFIELD is a non-parametric functional enrichment analysis approach described in the paper GARFIELD: GWAS analysis of regulatory or functional information enrichment with LD correction. Briefly, it is a method that leverages GWAS findings with regulatory or functional annotations (primarily from ENCODE and Roadmap epigenomics data) to find features relevant to a phenotype of interest. It performs greedy pruning of GWAS SNPs (LD r2 > 0.1) and then annotates them based on functional information overlap. Next, it quantifies Fold Enrichment (FE) at various GWAS significance cutoffs and assesses them by permutation testing, while matching for minor allele frequency, distance to nearest transcription start site and number of LD proxies (r2 > 0.8).

**biocViews** Software, StatisticalMethod, Annotation, FunctionalPrediction, GenomeAnnotation

License GPL-3

**NeedsCompilation** yes

VignetteBuilder knitr

Suggests knitr

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2 garfield-package

# **Contents**

	garfield-package .																										
	garfield.plot garfield.plot.fnx .																										
	garfield.run																										
Index																										11	L
garf	ield-package		AR ırio					-	,	Reg	gul	at	or	y c	or.	$F\iota$	ıno	cti	on	al	! In	nfe	or.	mo	ati	on	-

## **Description**

GARFIELD leverages GWAS findings with regulatory or functional annotations to find features relevant to a phenotype of interest. It performs greedy pruning of GWAS SNPs (LD r2>0.1) and then annotates them based on functional information overlap. Next, it quantifies Fold Enrichment (FE) at various GWAS significance cut-offs and assesses them by permutation testing, while matching for minor allele frequency, distance to nearest transcription start site and number of LD proxies (r2>0.8). Finally, it includes visualization tools.

## **Details**

Package: garfield Type: Package Version: 1.0

Date: 2015-12-14 License: GPL-3

See garfield.run for example analysis usage and garfield.plot for plotting examples.

### Author(s)

Sandro Morganella <email: sm22@sanger.ac.uk>

Maintainer: Valentina Iotchkova <email: vi1@sanger.ac.uk>

## References

Valentina Iotchkova, Graham Ritchie, Matthias Geihs, Sandro Morganella, Josine Min, Klaudia Walter, Nicholas Timpson, UK10K Consortium, Ian Dunham, Ewan Birney and Nicole Soranzo. GARFIELD - GWAS Analysis of Regulatory or Functional Information Enrichment with LD correction. In preparation

## See Also

garfield.run, garfield.plot

garfield.plot 3

garfield.plot Garfield plotting function
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## **Description**

garfield.plot is used for visualization of the enrichment analysis results obtained by the garfield.run permutation step. Internally, it uses garfield.plot.fnx which has been adapted from the 'radial.plot' function from the 'plotrix' package.

## Usage

```
garfield.plot(input_file, num_perm = 100000, output_prefix = "plot",
    plot_title = "", filter = 10, tr = Inf)
```

## **Arguments**

Input file name as produced by garfield.run permutation step. input\_file Number of permutations used in the garfield.run permutation step. num\_perm Figure file prefix. This would create the following files output prefix. Chromatin States.pdf, output\_prefix output\_prefix.Footprints.pdf, output\_prefix.Histone\_Modifications.pdf, output\_prefix.Peaks.pdf, output\_prefix.FAIRE.pdf, output\_prefix.Genic.pdf, output\_prefix.Hotspots.pdf and output\_prefix.TFBS.pdf Optional figure title plot\_title filter Optional filter for the minimum number of variants at a given threshold. Minimum of 1 should be used, but advisable to set to a larger value (e.g. 10). Threshold for denoting significance of an observed enrichment on a -log10 scale. tr A value of Inf denotes using default threshold of -log10(0.05/498).

## **Details**

This function is used for visualization of the enrichment analysis results and produces pdf figures for each class of annotations. Each figure shows the FE values (radial values) at different GWAS thresholds (bottom legend) for each annotation (outer circle and right legend). It further shows the significance at the top 4 GWAS thresholds (if present) as dots on the outer circle, with the most stringent threshold being shown at the inner most side.

## Value

No value is produced, instead output files are generated. See Details and 'output\_prefix' for more information.

# Author(s)

Sandro Morganella <email: sm22@sanger.ac.uk>

Maintainer: Valentina Iotchkova <email: vi1@sanger.ac.uk>

4 garfield.plot.fnx

## References

Valentina Iotchkova, Graham Ritchie, Matthias Geihs, Sandro Morganella, Josine Min, Klaudia Walter, Nicholas Timpson, UK10K Consortium, Ian Dunham, Ewan Birney and Nicole Soranzo. GARFIELD - GWAS Analysis of Regulatory or Functional Information Enrichment with LD correction. In preparation

#### See Also

```
garfield.run, garfield, garfield.plot.fnx
```

## **Examples**

```
garfield.run("tmp", data.dir=system.file("extdata",package = "garfield"),
    trait="trait",run.option = "prep", chrs = c(22),
    exclude = c(895, 975, 976, 977, 978, 979, 98))

garfield.run("tmp", data.dir=system.file("extdata",package = "garfield"),
    trait="", run.option = "perm", nperm = 1000,
    thresh = c(0.001, 1e-04, 1e-05), pt_thresh = c(1e-04, 1e-05),
    maf.bins = 2, tags.bins = 3, tss.bins = 3, prep.file = "tmp.prep",
    optim_mode = TRUE, minit = 100, thresh_perm = 0.05)

garfield.plot("tmp.perm", num_perm = 1000, output_prefix = "tmp",
    plot_title = "Sample run", filter = 1, tr = -log10(0.05))
#system("ls -lh tmp.*.pdf")
```

garfield.plot.fnx

Internal radial plotting function for the garfield.plot function

## **Description**

This function has been adapted from the 'radial.plot' function from the 'plotrix' package.

# Usage

```
garfield.plot.fnx(lengths, radial.pos = NULL, labels = NA, breaks = NA,
    label.pos = NULL, radlab = FALSE, start = 0, clockwise = FALSE,
    rp.type = "r", label.prop = 1.05, main = "", xlab = "", ylab = "",
    line.col = par("fg"), lty = par("lty"), lwd = par("lwd"),
    mar = c(2, 2, 3, 2), show.grid = TRUE, show.grid.labels = 4,
    show.radial.grid = TRUE, grid.col = "grey", grid.bg = "transparent",
    grid.left = FALSE, grid.unit = NULL, point.symbols = 1,
    point.col = par("fg"), show.centroid = FALSE, radial.lim = NULL,
    radial.labels = NULL, poly.col = NA, add = FALSE, ann.col = 1,
    ann.pch = 15, ann.col.mx = 1, compact = TRUE, ...)
```

garfield.plot.fnx 5

# Arguments

show.centroid

radial.labels

radial.lim

Whether to display a centroid.

lengths A numeric data vector or matrix. If 'lengths' is a matrix, the rows will be considered separate data vectors. A numeric vector or matrix of positions in radians. These are interpreted as radial.pos beginning at the right (0 radians) and moving counterclockwise. If 'radial.pos' is a matrix, the rows must correspond to rows of 'lengths'. labels Character strings to be placed at the outer ends of the lines. If set to NA, will suppress printing of labels, but if missing, the radial positions will be used. breaks A vector of (potentially different) labels to 'labels' according to which to draw radial lines. The positions of the labels around the plot in radians. label.pos radlab Whether to rotate the outer labels to a radial orientation. start Where to place the starting (zero) point. Defaults to the 3 o'clock position. Whether to interpret positive positions as clockwise from the starting point. The clockwise default is counterclockwise. rp.type Whether to draw (r)adial lines, a (p)olygon, (s)ymbols or some combination of these. If 'lengths' is a matrix and rp.type is a vector, each row of 'lengths' can be displayed differently. label.prop The label position radius as a proportion of the maximum line length. main The title for the plot. xlab, ylab Normally x and y axis labels are suppressed. line.col The color of the radial lines or polygons drawn. lty The line type(s) to be used for polygons or radial lines. lwd The line width(s) to be used for polygons or radial lines. Margins for the plot. Allows the user to leave space for legends, long labels, etc. mar Logical - whether to draw a circular grid. show.grid show.grid.labels Whether and where to display labels for the grid - see Details. show.radial.grid Whether to draw radial lines to the plot labels. grid.col Color of the circular grid. grid.bg Fill color of above. grid.left Whether to place the radial grid labels on the left side. grid.unit Optional unit description for the grid. The symbols for plotting (as in pch). point.symbols point.col Colors for the symbols.

The range of the grid circle. Defaults to 'pretty(range(lengths))', but if more

Optional labels for the radial grid. The default is the values of radial.lim.

than two values are passed, the exact values will be displayed.

garfield.plot.fnx

poly.col	Fill color if polygons are drawn. Use NA for no fill.
add	Whether to add one or more series to an existing plot.
ann.col	A vector of colours for symbols to be drawn on the outer circle.
ann.pch	A pch value for symbols to be drawn on the outer circle.
ann.col.mx	A matrix of colours for dots to be drawn on the outer circle, inside the symbols with 'ann.col' colours.
compact	A flag to specify if labels should be presented in a compact form or not.
	Additional arguments are passed to 'plot'.

#### **Details**

This function is only used internally as part of the garfield.plot function.

## Value

No value, but a plot is produced. See Details for further information.

## Author(s)

Sandro Morganella <email: sm22@sanger.ac.uk>

Maintainer: Valentina Iotchkova <email: vi1@sanger.ac.uk>

# References

Valentina Iotchkova, Graham Ritchie, Matthias Geihs, Sandro Morganella, Josine Min, Klaudia Walter, Nicholas Timpson, UK10K Consortium, Ian Dunham, Ewan Birney and Nicole Soranzo. GARFIELD - GWAS Analysis of Regulatory or Functional Information Enrichment with LD correction. In preparation

## See Also

```
garfield.plot, garfield.run, garfield
```

# **Examples**

```
DATA = rbind(rnorm(10,5,0.5),rnorm(10,3.5,0.5),rnorm(10,2,0.5))
garfield.plot.fnx(DATA,ann.col.mx=DATA!=0, ann.col=rep(1:2,each=5),
    ann.pch=15, rp.type="p",line.col=1:3,show.grid=TRUE, show.radial.grid=TRUE,
    labels=paste("label ",c(1:10)," ",sep=""),breaks=(1:10), radlab=TRUE,
    poly.col=1:3)
```

garfield.run GARFIELD enrichment analysis function	garfield.run	GARFIELD enrichment analysis function
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# Description

garfield. run is used to perform greedy pruning of variants from a genome-wide association study, calculate fold enrichment and test its significance at a given genome-wide significance threshold.

# Usage

```
garfield.run(out.file, data.dir, trait, run.option = "complete",
    chrs = c(1:22, "X"), exclude = c(895, 975, 976, 977, 978, 979, 98),
    nperm = 100000, thresh = c(0.1, 0.01, 0.001, 1e-04, 1e-05, 1e-06,
    1e-07, 1e-08), pt_thresh = c(1e-05, 1e-06, 1e-07, 1e-08),
    maf.bins = 5, tags.bins = 5, tss.bins = 5, prep.file = "",
    optim_mode = 1, minit = 100, thresh_perm = 1e-04)
```

## **Arguments**

out.file	Prefix for output file. Full garfield analysis creates out.file.prep and out.file.perm files from prunning and annotation step and permutation for significance testing step, respectively. These steps can additionally be run separately - for more information see run.option flag.
data.dir	Path to annotation and p-value files. The directory must contain "annotation", "maftssd", "pval" and "tags" subdirectories with per chromosome files of input data. See details section for further information.
trait	GWAS phenotype name. This must match a folder name in the data.dir folder. See details for more information.
run.option	an object specifying which part of the analysis to run. Valid options are complete, prep and perm, where prep denotes the preparation step (pruning and annotation of variants), perm denotes the permutation step (calculating fold enrichment and its significance) and complete executes both the preparation and permutation steps.
chrs	A vector of the chromosomes for which to run the enrichment analysis. chrs can have all or subsets of values from c(1:22, 'X').
exclude	A numeric vector of indeces of annotations for which LD tags should not be used for annotation of pruned variants. Value of -1 denotes using LD for annotation for all features.
nperm	A numeric value of the number of permutations to be performed.
thresh	A numeric vector of genome-wide significance thresholds to be used for fold enrichment calculation.
pt_thresh	A numeric vector of genome-wide significance thresholds to be used for calculating the significance of the observed fold enrichment. All values must be contained in the thresh vector.

maf.bins	A numeric value denoting the number of bins for the minor allele frequency matching during permutation testing. Must be greater or equal to 1.
tags.bins	A numeric value denoting the number of bins for the number of LD tags ( $r2>=0.8$ ) matching during permutation testing. Must be greater or equal to 1.
tss.bins	A numeric value denoting the number of bins for the distance to nearest transcription start site matching during permutation testing. Must be greater or equal to 1.
prep.file	File from pruning stage of algorithm. Only required if using the 'run.option'=perm flag.
optim_mode	A binary flag denoting whether to run fast version of method (1) or general version (0), where the fast version checks if significance of a given enrichment would still be possible to be obtained after 'minit' number of iterations and terminated permutations if not.
minit	An integer value for the minimum number of permutations to be performed before checking if thresh_perm condition can still be met. Only used if 'optim_mode'=1.
thresh_perm	After 'minit' number of permutations, at each iteration check if EmpPval can still reach 'thresh_perm' value. If not terminate permutations and output obtained results at that stage. Only used if 'optim_mode'=1.

#### **Details**

Output files: out.file.prep contains the genomic positions of pruned variants, p-values for association with the trait of interest, number of LD tags (r2>0.8), MAF, distance to the nearest TSS and binary representation of annotation information (with LD-tagging r2>0.8). out.file.perm contains enrichment analysis results for each annotation, where PThresh is the GWAS p-value threshold used for analysis, FE denotes the fold enrichment statistic (equals -1 if no sufficient data was available for the FE calculation), EmpPval shows the empirical p-value of enrichment (equals -1 if FE is calculated but significance of enrichment analysis is not run at that threshold), NAnnotThresh the number of variants at the threshold which are annotated with the given feature, NAnnot - the total number of annotated variants, NThresh - the total number of variants at that threshold and N - the total number of pruned variants. The remaining columns show additional information on the annotations used for analysis.

Data directory: data.dir, should point to a location containing "annotation", "maftssd", "pval" and "tags" subdirectories, where (i) the "pval" folder contains subfolders with trait names, which in turn contain per chromosome space separated files with genomic position in the first column and pvalue in the second. They should be named chr1, chr2, etc, and be numerically sorted with respect to genomic position; (ii) "annotation" should contain per chromosome space separated files with position in the first column and annotations in stacked binary format in the second. The files should be named chr1, chr2, etc and be sorted numerically according to position. Additionally the directory should contain a link\_file.txt file that links the annotations to relevant information about them; (iii) "maftssd" should contain per chromosome space separated files with position in the first column, minor allele frequency in the second and distance to the nearest TSS in the third. The files should be named chr1, chr2, etc and be sorted numerically according to position; (iv) "tags" should contain two subfolders named r01 and r08, which in turn contain per chromosome space separated files with variant position in the first column and comma separated positions of all variants with r2>0.1 or 0.8, respectively with the variant in the first column. The files should again be named chr1, chr2, etc

and be sorted numerically according to position. For further information on the data.dir structure, please see http://www.ebi.ac.uk/birney-srv/GARFIELD/documentation/GARFIELD.pdf

Pre-computed LD (European samples - UK10K sequence data), MAF, TSS distance, p-value files for two example traits (Crohn's Disease from the IBD Consortium and Height from the GIANT consortium) and annotation files for 1005 GENCODE, ENCODE and Roadmap Epigenomics annotations can be downloaded from http://www.ebi.ac.uk/birney-srv/GARFIELD/package/garfield-data.tar.gz. Note the data is 5.9Gb in compressed format and needs to be uncompressed prior to analysis (83Gb). Variant genomic position (build 37) is used as an identifier in all data files.

## Value

No value is produced, instead output files are generated. See Details and 'out.file' for more information.

## Author(s)

Sandro Morganella <email: sm22@sanger.ac.uk>

Maintainer: Valentina Iotchkova <email: vi1@sanger.ac.uk>

#### References

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## See Also

```
garfield.plot, garfield
```

## **Examples**

```
garfield.run("tmp", data.dir=system.file("extdata",package = "garfield"),
    trait="trait",run.option = "prep", chrs = c(22),
    exclude = c(895, 975, 976, 977, 978, 979, 98))

garfield.run("tmp", data.dir=system.file("extdata",package = "garfield"),
    run.option = "perm", nperm = 1000, thresh = c(0.001, 1e-04, 1e-05),
    pt_thresh = c(1e-04, 1e-05), maf.bins = 2, tags.bins = 3, tss.bins = 3,
    prep.file = "tmp.prep", optim_mode = TRUE, minit = 100, thresh_perm = 0.05)

if (file.exists("tmp.perm")){
    perm = read.table("tmp.perm", header=TRUE)
    head(perm)
} else { print("Error: tmp.perm does not exist!") }

###### To get the sample data for enrichment analysis in European samples
###### execute the following - note this can take a long time to run and
###### needs a substantial disk space (see Details)
####### needs a substantial disk space (see Details)
```

```
# download data and decompress
# system("wget http://www.ebi.ac.uk/birney-srv/GARFIELD/package/
# garfield-data.tar.gz")
# system("tar -zxvf garfield-data.tar.gz")
# if downloaded in current working directory use the following to execute
# garfield, otherwise please change data.dir location
# garfield.run("cd-meta.output", data.dir="garfield-data", trait="cd-meta",
\# run.option = "prep", chrs = c(1:22), exclude = c(895, 975, 976, 977, 978,
# 979, 980))
# garfield.run("cd-meta.output", data.dir="garfield-data", run.option = "perm",
\# nperm = 100000, thresh = c(0.1, 0.01, 0.001, 1e-04, 1e-05, 1e-06, 1e-07, 1e-08),
\# pt_thresh = c(1e-05, 1e-06, 1e-07, 1e-08), maf.bins = 5, tags.bins = 5,
# tss.bins = 5, prep.file = "cd-meta.output.prep", optim_mode = TRUE,
# minit = 100, thresh_perm = 0.0001)
# garfield.plot("cd-meta.output.perm", num_perm = 100000,
# output_prefix = "cd-meta.output", plot_title = "Crohn's Disease",
# filter = 10, tr = -\log 10(0.05/498))
```

# **Index**

```
* package
garfield-package, 2
garfield, 4, 6, 9
garfield (garfield-package), 2
garfield-package, 2
garfield.plot, 3, 6, 9
garfield.plot.fnx, 4, 4
garfield.run, 4, 6, 7
garfield_perm (garfield.run), 7
garfield_prep (garfield.run), 7
```