

# Package ‘AUCell’

April 15, 2020

**Type** Package

**Title** AUCell: Analysis of 'gene set' activity in single-cell RNA-seq data (e.g. identify cells with specific gene signatures)

**Version** 1.8.0

**Date** 2019-10-26

**Author** Sara Aibar, Stein Aerts. Laboratory of Computational Biology.  
VIB-KU Leuven Center for Brain & Disease Research. Leuven, Belgium.

**Maintainer** Sara Aibar <sara.aibar@kuleuven.vib.be>

**Description** AUCell allows to identify cells with active gene sets (e.g. signatures, gene modules...) in single-cell RNA-seq data. AUCell uses the "Area Under the Curve" (AUC) to calculate whether a critical subset of the input gene set is enriched within the expressed genes for each cell. The distribution of AUC scores across all the cells allows exploring the relative expression of the signature. Since the scoring method is ranking-based, AUCell is independent of the gene expression units and the normalization procedure. In addition, since the cells are evaluated individually, it can easily be applied to bigger datasets, subsetting the expression matrix if needed.

**URL** <http://scenic.aertslab.org>

**Imports** data.table, graphics, grDevices, GSEABase, methods, mixtools, R.utils, shiny, stats, SummarizedExperiment, BiocGenerics, S4Vectors, utils

**Enhances** doMC, doRNG, doParallel, foreach

**Suggests** Biobase, BiocStyle, devtools, dynamicTreeCut, DT, GEOquery, knitr, NMF, plotly, R2HTML, rbokeh, rmarkdown, Rtsne, testthat, zoo

**License** GPL-3

**biocViews** SingleCell, GeneSetEnrichment, Transcriptomics, Transcription, GeneExpression, WorkflowStep, Normalization

**LazyData** TRUE

**VignetteBuilder** knitr

**RoxygenNote** 6.1.1

**git\_url** <https://git.bioconductor.org/packages/AUCell>

**git\_branch** RELEASE\_3\_10

**git\_last\_commit** b1f460c

**git\_last\_commit\_date** 2019-10-29

**Date/Publication** 2020-04-14

## R topics documented:

aucellResults-class . . . . .	2
AUCCell_buildRankings . . . . .	4
AUCCell_calcAUC . . . . .	5
AUCCell_createViewerApp . . . . .	8
AUCCell_exploreThresholds . . . . .	9
AUCCell_plotHist . . . . .	12
AUCCell_plotTSNE . . . . .	13
nGenes . . . . .	15
orderAUC . . . . .	16
plotGeneCount . . . . .	16
plotTsne_cellProps . . . . .	17
updateAucellResults . . . . .	18
<b>Index</b>	<b>20</b>

---

aucellResults-class      *Wrapper to the matrix that stores the AUC or the cell rankings.*

---

### Description

This class extends SummarizedExperiment to contain the AUC matrix and cell rankings (as 'assays').

The results are stored in the assays slot, but they can be accessed through the regular methods (i.e. nrow, rownames... )

Types:

- "AUC": The assays contains the AUC for the gene-sets (or region-sets) & cells.

- "ranking": The assays contains the gene rankings for each cell.

### Usage

```
## S4 method for signature 'aucellResults'
show(object)
```

```
getAUC(object)
```

```
## S4 method for signature 'aucellResults'
getAUC(object)
```

```
getRanking(object)
```

```
## S4 method for signature 'aucellResults'
getRanking(object)
```

```
## S4 method for signature 'aucellResults'
cbind(..., deparse.level = 1)
```

**Arguments**

object            Results from AUCell\_buildRanking  
 ...                (Only for cbind)  
                   or AUCell\_calcAUC.  
 deparse.level    (Only for cbind)

**Value**

- show: Prints a summary of the object
- getAUC: Returns the matrix containing the AUC
- getRanking: Returns the matrix containing the rankings
- cbind: Combines objects by columns (cbind on assays); other other slots are conserved from the first object provided as argument. Both, ranking and AUC are calculated by column (cell or sample). Therefore, it is fine to merge objects as long as they come from equivalent datasets (and keep same genes/genesets, etc...)

**Examples**

```
# This example is run using a fake expression matrix.
# Therefore, the output will be meaningless.

##### Fake run of AUCell #####
set.seed(123)
exprMatrix <- matrix(data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
                     nrow=20,
                     dimnames=list(paste("Gene", 1:20, sep=""),
                                    paste("Cell", 1:500, sep="")))
dim(exprMatrix)

# Running AUCell
cells_rankings <- AUCell_buildRankings(exprMatrix)
fewGenes <- sample(rownames(exprMatrix), 10)
otherGenes <- sample(rownames(exprMatrix), 5)
geneSets <- list(geneSet1=fewGenes,
                geneSet2=otherGenes)
cells_AUC <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5, nCores=1)
#####

#Exploring the output:
cells_AUC

class(cells_AUC)

# Extracting the AUC matrix:
getAUC(cells_AUC)[,1:5]

# Subsetting and regular manipulation methods are also available:
cells_AUC[1:2,]
cells_AUC[,3:4]

dim(cells_AUC)
nrow(cells_AUC)
ncol(cells_AUC)
```

```

colnames(cells_AUC)
rownames(cells_AUC)

### Merging 2 objects (ranking or AUC):
sample1 <- cells_AUC[,10:20]
sample2 <- cells_AUC[,100:140]
cbind(sample1, sample2)

```

---

AUCell\_buildRankings *Build gene expression rankings for each cell*

---

### Description

Builds the "rankings" for each cell: expression-based ranking for all the genes in each cell.

The genes with same expression value are shuffled. Therefore, genes with expression '0' are randomly sorted at the end of the ranking.

These "rankings" can be seen as a new representation of the original dataset. Once they are calculated, they can be saved for future analyses.

### Usage

```

AUCell_buildRankings(exprMat, plotStats = TRUE, nCores = 1,
  verbose = TRUE, ...)

## S4 method for signature 'matrix'
AUCell_buildRankings(exprMat, plotStats = TRUE,
  nCores = 1, verbose = TRUE)

## S4 method for signature 'dgCMatrx'
AUCell_buildRankings(exprMat, plotStats = TRUE,
  nCores = 1, verbose = TRUE)

## S4 method for signature 'SummarizedExperiment'
AUCell_buildRankings(exprMat,
  plotStats = TRUE, nCores = 1, verbose = TRUE, assayName = NULL)

## S4 method for signature 'ExpressionSet'
AUCell_buildRankings(exprMat, plotStats = TRUE,
  nCores = 1, verbose = TRUE)

```

### Arguments

<code>exprMat</code>	<p>Expression matrix (genes as rows, cells as columns) The expression matrix can also be provided as one of the Bioconductor classes:</p> <ul style="list-style-type: none"> <li>• <a href="#">RangedSummarizedExperiment</a> and derived classes (e.g. <a href="#">SingleCellExperiment</a>): The matrix will be obtained through <code>assay(exprMatrix)</code>, -which will extract the first assay (usually the counts)- or the assay name given in <code>'assayName'</code></li> <li>• <a href="#">dgCMatrx-class</a>: Sparse matrix</li> <li>• <code>ExpressionSet</code>: The matrix will be obtained through <code>exprs(exprMatrix)</code></li> </ul>
----------------------	--

plotStats	Should the function plot the expression boxplots/histograms? (TRUE / FALSE). These plots can also be produced with the function <a href="#">plotGeneCount</a> .
nCores	Number of cores to use for computation.
verbose	Should the function show progress messages? (TRUE / FALSE)
...	Other arguments
assayName	Name of the assay containing the expression matrix (e.g. in <a href="#">SingleCellExperiment</a> objects)

### Details

It is important to check that most cells have at least the number of expressed/detected genes that are going to be used to calculate the AUC ('aucMaxRank' in 'calcAUC()'). The histogram provided by 'AUCell\_buildRankings()' allows to quickly check this distribution. 'plotGeneCount(exprMatrix)' allows to obtain only the plot before building the rankings.

### Value

data.table of genes (row) by cells (columns) with the ranking of the gene within the cell.

### See Also

Next step in the workflow: [AUCell\\_calcAUC](#).

See the package vignette for examples and more details: `vignette("AUCell")`

### Examples

```
# This example is run using a fake expression matrix.
# Therefore, the output will be meaningless.

##### Fake expression matrix #####
set.seed(123)
exprMatrix <- matrix(data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
                     nrow=20,
                     dimnames=list(paste("Gene", 1:20, sep=""),
                                   paste("Cell", 1:500, sep="")))
#####

cells_rankings <- AUCell_buildRankings(exprMatrix, plotStats=TRUE)
cells_rankings
```

---

AUCell\_calcAUC

*Calculate AUC*

---

### Description

Calculates the 'AUC' for each gene-set in each cell.

**Usage**

```

AUCell_calcAUC(geneSets, rankings, nCores = 1, normAUC = TRUE,
  aucMaxRank = ceiling(0.05 * nrow(rankings)), verbose = TRUE)

## S4 method for signature 'list'
AUCell_calcAUC(geneSets, rankings, nCores = 1,
  normAUC = TRUE, aucMaxRank = ceiling(0.05 * nrow(rankings)),
  verbose = TRUE)

## S4 method for signature 'character'
AUCell_calcAUC(geneSets, rankings, nCores = 1,
  normAUC = TRUE, aucMaxRank = ceiling(0.05 * nrow(rankings)),
  verbose = TRUE)

## S4 method for signature 'GeneSet'
AUCell_calcAUC(geneSets, rankings, nCores = 1,
  normAUC = TRUE, aucMaxRank = ceiling(0.05 * nrow(rankings)),
  verbose = TRUE)

## S4 method for signature 'GeneSetCollection'
AUCell_calcAUC(geneSets, rankings,
  nCores = 1, normAUC = TRUE, aucMaxRank = ceiling(0.05 *
  nrow(rankings)), verbose = TRUE)

```

**Arguments**

geneSets	List of gene-sets (or signatures) to test in the cells. The gene-sets should be provided as <a href="#">GeneSet</a> , <a href="#">GeneSetCollection</a> or character list (see examples).
rankings	'Rankings' created for this dataset with <a href="#">AUCell_buildRankings</a> .
nCores	Number of cores to use for computation.
normAUC	Wether to normalize the maximum possible AUC to 1 (Default: TRUE).
aucMaxRank	Threshold to calculate the AUC (see 'details' section)
verbose	Should the function show progress messages? (TRUE / FALSE)

**Details**

In a simplified way, the AUC value represents the fraction of genes, within the top X genes in the ranking, that are included in the signature. The parameter 'aucMaxRank' allows to modify the number of genes (maximum ranking) that is used to perform this computation. By default, it is set to 5% of the total number of genes in the rankings. Common values may range from 1 to 20%.

**Value**

Matrix with the AUC values (gene-sets as rows, cells as columns).

**See Also**

Previous step in the workflow: [AUCell\\_buildRankings](#). Next step in the workflow: [AUCell\\_exploreThresholds](#).

See the package vignette for examples and more details: `vignette("AUCell")`

**Examples**

```

# This example is run using a fake expression matrix.
# Therefore, the output will be meaningless.

##### Fake expression matrix #####
set.seed(123)
exprMatrix <- matrix(data=sample(c(rep(0, 5000)), sample(1:3, 5000, replace=TRUE)),
                    nrow=20,
                    dimnames=list(paste("Gene", 1:20, sep=""),
                                   paste("Cell", 1:500, sep="")))
#####

##### Previous step in the workflow #####
# Step 1.
cells_rankings <- AUCCell_buildRankings(exprMatrix)
#####

##### Step 2: Calculate AUC #####

# In this example we use two gene sets: 10 and 5 random genes
# (see other formatting examples at the end)
fewGenes <- sample(rownames(exprMatrix), 10)
otherGenes <- sample(rownames(exprMatrix), 5)

geneSets <- list(geneSet1=fewGenes,
                 geneSet2=otherGenes)
geneSets

# Calculate AUC with the rankings from Step 1.
# To be able to run this fake example (which contain only 20 genes),
# we use aucMaxRank=5 (top 25% of the genes in the ranking)
set.seed(123)
cells_AUC <- AUCCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5, nCores=1)

# Format of the output:
cells_AUC

# To subset & access the AUC slot (as matrix):
cells_AUC[1:2,]
cells_AUC[,3:4]
getAUC(cells_AUC)[,1:5]

# These methods are also available:
dim(cells_AUC)
nrow(cells_AUC)
ncol(cells_AUC)
colnames(cells_AUC)
rownames(cells_AUC)

#####
# Alternatives for the input of gene sets:

# a) Character vector (i.e. only one gene-set)
# It will take the default name 'geneSet'
fewGenes

```

```

test <- AUCell_calcAUC(fewGenes, cells_rankings, aucMaxRank=5)

# b) List
geneSets <- list(geneSet1=fewGenes,
                 geneSet2=otherGenes)
geneSets
test <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5)

# c) GeneSet object (from GSEABase)
library(GSEABase)
geneSetOne <- GeneSet(fewGenes, setName="geneSetOne")
geneSetOne
test <- AUCell_calcAUC(geneSetOne, cells_rankings, aucMaxRank=5)

# d) GeneSetCollection object (from GSEABase)
geneSetTwo <- GeneSet(otherGenes, setName="geneSetTwo")
geneSets <- GeneSetCollection(geneSetOne, geneSetTwo)
geneSets
test <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5)

```

---

AUCell\_createViewerApp

*Create AUCell viewer app*


---

## Description

Creates a Shiny app to explore AUCell results

## Usage

```
AUCell_createViewerApp(auc, thresholds = NULL, tSNE = NULL,
  exprMat = NULL, cellInfo = NULL, colVars = NULL)
```

## Arguments

auc	AUC object returned by <a href="#">AUCell_calcAUC</a>
thresholds	Thresholds corresponding to each gene set (optional)
tSNE	t-SNE coordinates for the cells (optional). The row names should correspond to the cell ID. The column names should be "tsne1" and "tsne2".
exprMat	Expression matrix (optional)
cellInfo	Phenodata (optional)
colVars	Color for the phenodata variables (as list, optional)

## Value

Thresholds and cells selected within the app (as list).

## Note

With lasso: "To make a multiple selection, press the SHIFT key. To clear the selection, press the ESC key."



**Examples**

```
#####
# Fake run of AUCell
set.seed(123)
exprMatrix <- matrix(
  data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
  nrow=20,
  dimnames=list(paste("Gene", 1:20, sep=""),
                paste("Cell", 1:500, sep="")))
geneSets <- list(geneSet1=sample(rownames(exprMatrix), 10),
                 geneSet2=sample(rownames(exprMatrix), 5))

cells_rankings <- AUCell_buildRankings(exprMatrix)
cells_AUC <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5, nCores=1)
selectedThresholds <- NULL
cellsTsne<- Rtsne::Rtsne(t(exprMatrix),max_iter = 10)$Y
rownames(cellsTsne) <- colnames(exprMatrix)

cellInfo <- data.frame(cellType1=sample(LETTERS[1:3],ncol(exprMatrix), replace=TRUE),
                       cellType2=sample(letters[5:7],ncol(exprMatrix), replace=TRUE),
                       nGenes=abs(rnorm(ncol(exprMatrix))),
                       row.names=colnames(exprMatrix))

#####

library(shiny); library(rbokeh)

# Create app
aucellApp <- AUCell_createViewerApp(auc=cells_AUC, thresholds=selectedThresholds,
                                   tSNE=cellsTsne, exprMat=exprMatrix, cellInfo=cellInfo)

# The exact commands to launch the app depend on the R settings
# For example:
# options(shiny.host="0.0.0.0")
# savedSelections <- runApp(aucellApp)
# (see Shiny's doc for help)
```

---

AUCell\_exploreThresholds

*AUCell\_exploreThresholds*


---

**Description**

Plots all the AUC histograms (per gene-set) and calculates several likely thresholds for each gene-set

**Usage**

```
AUCell_exploreThresholds(cellsAUC, thrP = 0.01, nCores = 1,
  smallestPopPercent = 0.25, plotHist = TRUE, densAdjust = 2,
  assignCells = FALSE, nBreaks = 100, verbose = TRUE)
```

```
getThresholdSelected(aucellThresholds)
```

```
getAssignments(aucellThresholds)
```

### Arguments

cellsAUC	AUC object returned by <a href="#">AUCell_calcAUC</a> .
thrP	Probability to determine outliers in some of the distributions (see 'details' section). By default it is set to 1% (thrP): if there are 3000 cells in the dataset, it is expected that approximately 30 cells are over this threshold if the AUC is normally distributed.
nCores	Number of cores to use for computation.
smallestPopPercent	Size (percentage) of the smallest population of cells expected. Used to calculate some of the thresholds.
plotHist	Whether to plot the AUC histograms. (TRUE / FALSE)
densAdjust	Parameter for the density curve. (See <code>density</code> for details).
assignCells	Return the list of cells that pass the automatically selected threshold? (TRUE/FALSE)
nBreaks	Number of bars to plot in the histograms.
verbose	Should the function show progress messages? (TRUE / FALSE)
aucellThresholds	For aux functions: Output from <code>AUCell_exploreThresholds</code>

### Details

To ease the selection of an assignment threshold, this function adjusts the AUCs of each gene-set to several distributions and calculates possible thresholds:

- `minimumDens` (plot in Blue): Inflection point of the density curve. This is usually a good option for the ideal situation with bimodal distributions.  
To avoid false positives, by default this threshold will not be chosen if the second distribution is higher (i.e. the majority of cells have the gene-set "active").
- `L_k2` (plot in Red): Left distribution, after adjusting the AUC to a mixture of two distributions. The threshold is set to the right (prob:  $1 - (\text{thrP}/n\text{Cells})$ ). Only available if 'mixtools' package is installed.
- `R_k3` (plot in Pink): Right distribution, after adjusting the AUC to a mixture of three distributions. The threshold is set to the left (prob: `thrP`). Only available if 'mixtools' package is installed.
- `Global_k1` (plot in Grey): "global" distribution (i.e. mean and standard deviations of all cells). The threshold is set to the right (prob:  $1 - (\text{thrP}/n\text{Cells})$ ).  
The threshold based on the global distribution is ignored from the automatic selection unless the mixed models are overlapping.

Note: If `assignCells=TRUE`, the highest threshold is used to select cells. However, keep in mind that this function is only meant to ease the selection of the threshold, and we highly recommend to look at the AUC histograms and adjust the threshold manually if needed. We recommend to be specially aware on gene-sets with few genes (10-15) and thresholds that are set extremely low.

**Value**

List with the following elements for each gene-set:

- 'aucThr' Thresholds calculated with each method (see 'details' section), and the number of cells that would be assigned using that threshold.  
If assignCells=TRUE, the threshold selected automatically is the highest value (in most cases, excluding the global distribution).
- 'assignment' List of cells that pass the selected AUC threshold (if assignCells=TRUE)

If plotHist=TRUE the AUC histogram is also plot, including the distributions calculated and the corresponding thresholds in the same color (dashed vertical lines). The threshold that is automatically selected is shown as a thicker non-dashed vertical line.

**See Also**

Previous step in the workflow: [AUCell\\_calcAUC](#).

See the package vignette for examples and more details: `vignette("AUCell")`

**Examples**

```
# This example is run using a fake expression matrix.
# Therefore, the output will be meaningless.

##### Fake expression matrix #####
set.seed(123)
exprMatrix <- matrix(data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
                      nrow=20,
                      dimnames=list(paste("Gene", 1:20, sep=""),
                                    paste("Cell", 1:500, sep="")))

dim(exprMatrix)
#####

##### Previous steps in the workflow #####
# Step 1.
cells_rankings <- AUCell_buildRankings(exprMatrix, plotStats=FALSE)

# Step 2.
# (Gene sets: random genes)
geneSets <- list(geneSet1=sample(rownames(exprMatrix), 10),
                geneSet2=sample(rownames(exprMatrix), 5))
cells_AUC <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5)
#####

##### Step 3: Assign cells #####

# 1. Plot histograms and obtain some pre-computed thresholds
# (this example is only meant to show the interface/arguments of the function,
# see the vignette for meaningful examples)
set.seed(123)
par(mfrow=c(1,2)) # Plot is divided into one row and two columns
thresholds <- AUCell_exploreThresholds(cells_AUC, plotHist=TRUE)
thresholds$geneSet1$aucThr

# 2. Obtain cells over a given threshold:
names(which(getAUC(cells_AUC)["geneSet1",] > 0.19))
```

```
# Alternative: assign cells according to the 'automatic' threshold
cells_assignment <- AUCell_exploreThresholds(cells_AUC,
                                           plotHist=FALSE, assignCells=TRUE)

# Cells assigned:
getAssignments(cells_assignment)
# Threshold applied:
getThresholdSelected(cells_assignment)
```

---

AUCell\_plotHist      *Plot AUC histogram*

---

### Description

Plots the distribution of AUC across the cells (for each gene-set) as an histogram.

### Usage

```
AUCell_plotHist(cellsAUC, aucThr = max(cellsAUC), nBreaks = 100,
                onColor = "dodgerblue4", offColor = "slategray2", ...)
```

### Arguments

cellsAUC	Subset of the object returned by <a href="#">AUCell_calcAUC</a> (i.e. including only the gene-sets to plot)
aucThr	AUC value planned to use as threshold (to make sure the X axis includes it), if any. Otherwise, the X axis extends to cover only the AUC values plotted.
nBreaks	Number of 'bars' to plot (breaks argument for hist function).
onColor	Color for the bars that pass the AUC threshold
offColor	Color for the bars that do not pass the AUC threshold
...	Other arguments to pass to <a href="#">hist</a> function.

### Value

List of histogram objects (invisible).

### See Also

See the package vignette for examples and more details: `vignette("AUCell")`

### Examples

```
# This example is run using a fake expression matrix.
# Therefore, the output will be meaningless.

##### Fake expression matrix #####
set.seed(123)
exprMatrix <- matrix(data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
                    nrow=20,
                    dimnames=list(paste("Gene", 1:20, sep="")),
```

```

paste("Cell", 1:500, sep=""))
dim(exprMatrix)
#####

##### Beginning of the workflow #####
# Step 1.
cells_rankings <- AUCCell_buildRankings(exprMatrix, plotStats=FALSE)

# Step 2.
# (Gene set: 10 random genes)
genes <- sample(rownames(exprMatrix), 10)
geneSets <- list(geneSet1=genes)
# (aucMaxRank=5 to run with this fake example, it will return 'high' AUC values)
cells_AUC <- AUCCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5)
#####

# Plot histogram:
AUCCell_plotHist(cells_AUC["geneSet1",], nBreaks=10)

```

AUCCell\_plotTSNE

*Plot***Description**

Plots the AUC histogram and t-SNE coloured by AUC, binary activity and TF expression

**Usage**

```

AUCCell_plotTSNE(tSNE, exprMat = NULL, cellsAUC = NULL,
  thresholds = NULL, reorderGeneSets = FALSE, cex = 1, alphaOn = 1,
  alphaOff = 0.2, borderColor = adjustcolor("lightgray", alpha.f =
  0.1), offColor = "lightgray", plots = c("histogram", "binaryAUC",
  "AUC", "expression"), exprCols = c("goldenrod1", "darkorange",
  "brown"), asPNG = FALSE, ...)

```

**Arguments**

tSNE	t-SNE coordinates (e.g. tSNE\$Y)
exprMat	Expression matrix
cellsAUC	AUC (as returned by calcAUC)
thresholds	Thresholds returned by AUCCell
reorderGeneSets	Whether to reorder the gene sets based on AUC similarity
cex	Scaling factor for the dots in the scatterplot
alphaOn	Transparency for the dots representing "active" cells
alphaOff	Transparency for the dots representing "inactive" cells
borderColor	Border color for the dots (scatterplot)
offColor	Color for the dots representing "inactive" cells
plots	Which plots to generate? Select one or multiple: plots=c("histogram", "binaryAUC", "AUC", "exp

exprCols           Color scale for the expression  
 asPNG               Output each individual plot in a .png file? (can also be a directory)  
 ...                 Other arguments to pass to [hist](#) function.

### Details

To avoid calculating thresholds, set thresholds to FALSE

### Value

Returns invisible: cells\_trhAssignment

### See Also

List of vignettes included in the package: `vignette(package="AUCell")`

### Examples

```
#####
# Fake run of AUCell
set.seed(123)
exprMatrix <- matrix(
  data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
  nrow=20,
  dimnames=list(paste("Gene", 1:20, sep=""),
                paste("Cell", 1:500, sep="")))
geneSets <- list(geneSet1=sample(rownames(exprMatrix), 10),
                 geneSet2=sample(rownames(exprMatrix), 5))

cells_rankings <- AUCell_buildRankings(exprMatrix, plotStats = FALSE)
cells_AUC <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5, nCores=1)
selectedThresholds <- rowMeans(getAUC(cells_AUC))
cellsTsne <- Rtsne::Rtsne(t(exprMatrix), max_iter = 10)$Y
rownames(cellsTsne) <- colnames(exprMatrix)
#####

par(mfrow=c(2,3))
thrs <- AUCell_plotTSNE(tSNE=cellsTsne, exprMat=NULL,
                       cellsAUC=cells_AUC, thresholds=selectedThresholds,
                       plots = c("histogram", "binaryAUC", "AUC"))

#####
# Color based on the known phenodata:
cellInfo <- data.frame(cellType1=sample(LETTERS[1:3], ncol(exprMatrix), replace=TRUE),
                      cellType2=sample(letters[5:7], ncol(exprMatrix), replace=TRUE),
                      nGenes=abs(rnorm(ncol(exprMatrix))),
                      row.names=colnames(exprMatrix))
colVars <- list(cellType2=setNames(c("skyblue", "magenta", "darkorange"), letters[5:7]))
# dev.off()
plotTsne_cellProps(cellsTsne, cellInfo, colVars=colVars)
```

---

nGenes

*GeneSet methods*

---

## Description

Functions to manipulate GeneSet and GeneSetCollection objects (from package GSEABase)

## Usage

```
nGenes(geneSet)

## S4 method for signature 'GeneSet'
nGenes(geneSet)

## S4 method for signature 'GeneSetCollection'
nGenes(geneSet)

subsetGeneSets(geneSets, geneNames)

## S4 method for signature 'GeneSetCollection'
subsetGeneSets(geneSets, geneNames)

setGeneSetNames(geneSets, newNames)

## S4 method for signature 'GeneSetCollection'
setGeneSetNames(geneSets, newNames)
```

## Arguments

geneSet	One gene-set ( <a href="#">GeneSet</a> )
geneSets	Gene-set collection ( <a href="#">GeneSetCollection</a> )
geneNames	Gene names (for subset)
newNames	New names (to assign to the gene sets)

## Value

- **nGenes()**: provides the number of genes in the gene-set, or each of the gene-sets in a collection
- **subsetGeneSets()**: Subsets each of the gene-sets in a collection to contain only the genes in the given list. Equivalent to `intersect()`, but keeping the original gene-set name.
- **setGeneSetNames()**: Modifies the name of each gene-set in a collection

## Examples

```
library(GSEABase)
genes_1 <- GeneSet(paste("Gene", 1:20, sep=""), setName="geneSet1")
genes_2 <- GeneSet(paste("Gene", 18:22, sep=""), setName="geneSet2")
geneSets <- GeneSetCollection(genes_1, genes_2)

nGenes(genes_1)
```

```
nGenes(geneSets)

subsetGeneSets(geneSets, paste("Gene", 15:20, sep=""))

geneSets_newNames <- setGeneSetNames(geneSets, c("one", "two"))
names(geneSets_newNames)
```

---

orderAUC

*orderAUC*


---

### Description

Reorder the gene-sets based on AUC similarity

### Usage

```
orderAUC(auc)
```

### Arguments

auc                    AUC (as returned by calcAUC)

### Value

gene-set names in the suggested order

### Examples

```
# cellsAUC <- cellsAUC[orderAUC(cellsAUC),]
```

---

plotGeneCount

*plotGeneCount*


---

### Description

Plots a histogram and boxplot for the number of genes detected in each cell.

### Usage

```
plotGeneCount(exprMat, plotStats = TRUE, verbose = TRUE)
```

### Arguments

exprMat                Expression matrix (genes as rows, cells as columns)

plotStats              Logical. If true, it plots the histogram, otherwise only calculates the percentages of genes detected.

verbose                Should the function show progress messages? (TRUE / FALSE)



**Details**

It is important to check that most cells have at least the number of expressed/detected genes that are going to be used to calculate the AUC ('aucMaxRank' in 'calcAUC()'). The histogram provided by 'AUCell\_buildRankings()' allows to quickly check this distribution. 'plotGeneCount(exprMatrix)' allows to obtain only the plot before building the rankings.

**Value**

Quantiles with the number of genes detected by cell (invisible). This result is also printed if verbose=TRUE.

**See Also**

See the package vignette for more details: vignette("AUCell")

**Examples**

```
### (Fake expression matrix)
exprMatrix <- matrix(sample(c(rep(0, 500), sample(1:3, 500, replace=TRUE))),
  nrow=20)
rownames(exprMatrix) <- paste("Gene", 1:20, sep="")
colnames(exprMatrix) <- paste("Sample", 1:50, sep="")
###

plotGeneCount(exprMatrix)
title(sub="Fake expression matrix")
```

---

plotTsne\_cellProps      *plotTsne\_cellProps*

---

**Description**

Plots the t-SNE coloured based on the known the cell properties

**Usage**

```
plotTsne_cellProps(tSNE, cellInfo, colVars = NULL, cex = 1, sub = "",
  gradientCols = c("yellow", "orange", "red"), showLegend = TRUE)
```

**Arguments**

tSNE	t-SNE coordinates (e.g. tSNE\$Y)
cellInfo	Dataframe with cell phenodata
colVars	Colors for the cell properties (optional)
cex	Scaling factor for the dots in the scatterplot
sub	Subtitle (e.g. tSNE type)
gradientCols	Gradient colors for numerical variables
showLegend	Whether to show the legend

**Value**

Plots the t-SNE

**Examples**

```
#####
# Fake run of AUCell
set.seed(123)
exprMatrix <- matrix(
  data=sample(c(rep(0, 5000), sample(1:3, 5000, replace=TRUE))),
  nrow=20,
  dimnames=list(paste("Gene", 1:20, sep=""),
                paste("Cell", 1:500, sep="")))
geneSets <- list(geneSet1=sample(rownames(exprMatrix), 10),
                 geneSet2=sample(rownames(exprMatrix), 5))

cells_rankings <- AUCell_buildRankings(exprMatrix, plotStats = FALSE)
cells_AUC <- AUCell_calcAUC(geneSets, cells_rankings, aucMaxRank=5, nCores=1)
selectedThresholds <- rowMeans(getAUC(cells_AUC))
cellsTsne <- Rtsne::Rtsne(t(exprMatrix), max_iter = 10)$Y
rownames(cellsTsne) <- colnames(exprMatrix)
#####

par(mfrow=c(2,3))
thrs <- AUCell_plotTSNE(tSNE=cellsTsne, exprMat=NULL,
                       cellsAUC=cells_AUC, thresholds=selectedThresholds,
                       plots = c("histogram", "binaryAUC", "AUC"))

#####
# Color based on the known phenodata:
cellInfo <- data.frame(cellType1=sample(LETTERS[1:3], ncol(exprMatrix), replace=TRUE),
                       cellType2=sample(letters[5:7], ncol(exprMatrix), replace=TRUE),
                       nGenes=abs(rnorm(ncol(exprMatrix))),
                       row.names=colnames(exprMatrix))
colVars <- list(cellType2=setNames(c("skyblue", "magenta", "darkorange"), letters[5:7]))
# dev.off()
plotTsne_cellProps(cellsTsne, cellInfo, colVars=colVars)
```

---

updateAucellResults     *Update AUCell results*

---

**Description**

Updates the AUC scores provided by AUCell from a previous version.

**Usage**

```
updateAucellResults(oldAucObject, objectType = "AUC")
```

**Arguments**

oldAucObject    Object to update  
objectType     Either "AUC" or "ranking" indicating the object type

**Value**

Updated version of the object as [aucellResults](#).

**Examples**

```
oldAuc <- matrix(data=1:2000, nrow=50, ncol=40)  
updateAucellResults(oldAuc)
```

# Index

AUCell\_buildRankings, [4, 6](#)  
AUCell\_buildRankings, dgCMatrx-method  
    (AUCell\_buildRankings), [4](#)  
AUCell\_buildRankings, ExpressionSet-method  
    (AUCell\_buildRankings), [4](#)  
AUCell\_buildRankings, matrix-method  
    (AUCell\_buildRankings), [4](#)  
AUCell\_buildRankings, SummarizedExperiment-method  
    (AUCell\_buildRankings), [4](#)  
AUCell\_calcAUC, [5, 5, 8, 10–12](#)  
AUCell\_calcAUC, character-method  
    (AUCell\_calcAUC), [5](#)  
AUCell\_calcAUC, GeneSet-method  
    (AUCell\_calcAUC), [5](#)  
AUCell\_calcAUC, GeneSetCollection-method  
    (AUCell\_calcAUC), [5](#)  
AUCell\_calcAUC, list-method  
    (AUCell\_calcAUC), [5](#)  
AUCell\_createViewerApp, [8](#)  
AUCell\_exploreThresholds, [6, 9](#)  
AUCell\_plotHist, [12](#)  
AUCell\_plotTSNE, [13](#)  
aucellResults, [19](#)  
aucellResults (aucellResults-class), [2](#)  
aucellResults-class, [2](#)  
  
cbind (aucellResults-class), [2](#)  
  
dgCMatrx-class, [4](#)  
  
GeneSet, [6, 15](#)  
GeneSetCollection, [6, 15](#)  
getAssignments  
    (AUCell\_exploreThresholds), [9](#)  
getAUC (aucellResults-class), [2](#)  
getAUC, aucellResults-method  
    (aucellResults-class), [2](#)  
getRanking (aucellResults-class), [2](#)  
getRanking, aucellResults-method  
    (aucellResults-class), [2](#)  
getThresholdSelected  
    (AUCell\_exploreThresholds), [9](#)  
  
hist, [12, 14](#)  
  
nGenes, [15](#)  
nGenes, GeneSet-method (nGenes), [15](#)  
nGenes, GeneSetCollection-method  
    (nGenes), [15](#)  
  
orderAUC, [16](#)  
  
plotGeneCount, [5, 16](#)  
plotTsne\_cellProps, [17](#)  
  
RangedSummarizedExperiment, [4](#)  
  
setGeneSetNames (nGenes), [15](#)  
setGeneSetNames, GeneSetCollection-method  
    (nGenes), [15](#)  
show (aucellResults-class), [2](#)  
show, aucellResults-method  
    (aucellResults-class), [2](#)  
SingleCellExperiment, [4, 5](#)  
subsetGeneSets (nGenes), [15](#)  
subsetGeneSets, GeneSetCollection-method  
    (nGenes), [15](#)  
  
updateAucellResults, [18](#)